

POTENTIAL OF BIOMETHANE FROM WASHED ASHORE ALGAE IN GULF OF RIGA

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Abstract. There is a strong tourism industry with a high recreational value in the southern Baltic, which is adversely affected by the algae that accumulate along the coast. The project's concept of transforming the problem into a resource by preventing eutrophication through biogas production has several benefits for the region: removal of phosphorus and nitrogen from the Baltic Sea and their return to arable land (phosphorus is a limited resource to be saved for future food production); improved coastal recreation values and, in the long term, the regional economy; reduction of greenhouse gas (GHG) emissions from algae. Algae can provide bioenergy not only from agricultural land, but from our seas and oceans. Seaweed can be used to purify nutrient-enriched water (associated with salmon, for example, on farms). Many species of seaweed can be classified in many ways, for instance, by color. Genetic difference between green seaweed *Ulva lactuca* and brown seaweed *Fucus* is quite large. *U. lactuca* has a high sulfur content and usually has a carbon to nitrogen (C: N) ratio of less than 10, which makes anaerobic fermentation difficult. To determine the potential of biogas from different seaweeds of the Gulf of Riga, a study was carried out using laboratory equipment. In the study, 16 different bioreactors fermented at 38 °C three different types of algae most common in the Gulf of Riga, taken from shore-washed seaweed piles. On average, fermentation for 32 days yielded 0.276 L·g⁻¹dom biogas (0.046 L·g⁻¹dom methane) from brown algae, and 0.248 L·g⁻¹dom biogas (0.027 L·g⁻¹dom methane) from red algae. An average of 0.425 L·g⁻¹dom biogas (0.071 L·g⁻¹dom methane) was obtained from green algae. The study shows that long-lived algae can be obtained in small amounts of methane per unit dry organic matter from land-littered ponds, if no measures are taken to condition them.

Keywords: biogas, methane, seaweed, anaerobic fermentation, digestate.

Introduction

The United Nations Intergovernmental Panel on Climate Change (IPCC) report [1], published September 25, 2019 in Monte Carlo, indicates that changes will continue and will be irreversible, even if the stabilization of climate is reached. Significantly reducing emissions, thus limiting the loss of glaciers and at the same time focusing on adaptation measures, it is possible to protect the millions of people's livelihoods and to provide a basis for sustainable development, ranging from mountain areas to coastal and sea regions. Since 80 years, the oceans have absorbed 20-30 % of the total anthropogenic emissions. Since 1993, ocean warming rate has more than doubled. Data sets covering the period of 1970-2010, show that the top (1000 m) ocean surface has lost oxygen levels of 0.5-3.3 %, which in the future will be affected by at least 60 % of all ocean area. The more CO₂ oceans absorb, the more acidification of ocean waters takes place. The ocean acidification process and the level of oxygen loss, changing ecosystem structures and their ability to function, have a direct impact on many species, such as corals, mussels and fish. Ocean warming has also led to a reduction in the total catch. In turn, in context of adaptation measures there will be urgent need to adapt not only to the sea level raise, but also higher waves. Wave height will increase significantly in the Southern Ocean, the Pacific Ocean to the east and the Baltic Sea [1] (IPCC report summary 26.09. 2019).

South Baltic area has a strong tourism industry with high recreational value of the negative impact from algae that builds up along the coast. However, the problem can be turned into opportunity by applying biogas production and at the same time preventing eutrophication of coastal waters. The region would have several benefits:

- phosphorus and nitrogen separation from the Baltic Sea and the return to arable land;
- reduction of organic matter leakage from agricultural lands by removing wetlands;
- increasing biodiversity;
- improvement of coastal water availability for juvenile fish and other important organisms.

Justification of bioenergy production from algae is addressed by debate on food versus fuel production in context of Land Use, Land-Use Change and Forestry (LULUCF). Using limited resources of arable land (0.2 ha of arable land per capita worldwide) raises ethical questions on energy production in competition with food production. Algae from seas and oceans can produce bioenergy

instead of using agricultural land for this purpose. Seaweed can purify waters rich in nutrients and can benefit salmon breeding, for instance, at coastal farms.

There may be a number of seaweed species classified for instance by color. The genetic difference between the green seaweed *Ulva lactuca* and brown seaweed *Fucus* is quite large. *U. lactuca* contains a lot of sulfur and has generally the carbon and nitrogen (C: N) ratio of less than 10, so the anaerobic fermentation is difficult. There are about 10,000 species of seaweed. Jard et al. (2013) [2] specify seaweeds in three broad parts of the following colors: brown, red and green algae.

- brown algae include *Saccharina latissima*, *Himantalia elongata*, *Laminaria digitata*, *Fucusserratus*, *Ascophyllum nodosum*, *Undaria pinnatifida*, *Sargassum muticum*;
- red seaweed include *Gracilaria verrucosa*, *Palmaria palmata* and *Asparagopsis armata*;
- green algae include *Codiumtomen tosum* and *Ulva lactuca*.

Biogas yield is dependent on the volatile solids (VS) or total solids (TS) raw material. Bruhnetal (2011) [3] reported about content 57 % of *Ulva lactuca* VS in Denmark. Allen and al. (2014) [4] found 58 % VS collected from *U. lactuca* content in June 2011 from an estuary in West Cork. Jard et al. (2013) [2] concluded that the summer algae accumulate more carbon and the C: N ratio would increase and therefore *Ulva* is with higher VS content.

Using the stoichiometric method Allenetal (2014) [4] showed that the *Ulva* theoretical maximum methane production could be 431 L·kg⁻¹VS with 51.5 % methane content. Allenetal (2014) [4] reported that algae collected at West Cork in 2013 shown C:N ratio exceeded 20. The optimum anaerobic fermentation C:N ratio is from 20 to 1 up to 30 to 1. Total VS were no more than 8 % up to 19 % from naturally wet weight.

Values of fresh *Ulva* of Ireland and Denmark are very similar [5-8]. Bruhn et al. (2011) [3] *U. lactuca* collected from Sedenbeach (Odense fjords), Denmark were the ratio VS/TS was very similar (57 % and 58 %). Raw, fresh *Ulva* collected in Ireland show 183 L CH₄·kg⁻¹VS, while *Ulva* in Denmark generated 174 L CH₄·kg⁻¹VS [8]. This suggests that similar results can be obtained from *Ulva* in the Northern part of Europe.

Biogas potential of red algae *Polysiphonia* was studied at the Linköping University, Biswas [9] got biogas 0.25-0.27 m³·kg⁻¹VS from untreated algae. Out of autoclave treated algae maximum 0.25 m³·kg⁻¹VS methane was obtained.

Seaweed is appropriate to use in mix with other feedstock. For example, in Denmark Solrod commune a biogas plant together with 53 200 t·y⁻¹ of pig slurry 7400 t·y⁻¹ seaweed were used. It is estimated that around 1,141 t·km⁻¹ of coast can be collected. From algae wet weight only 4 m³·t⁻¹ (from pig manure 10,7 m³·t⁻¹) can be obtained [10] (Solrød Kommune 2014). Seaweeds are also used in Sweden Trelleborg commune biogas plant Smyga. Although, algae collected on the coast had impurity of inorganics (by weight) up to 80 % [11] and for purification of sand special equipment was installed.

Biogas yield from the Gulf of Riga algae in Latvia so far has not been studied.

Materials, techniques and experiment description

As raw materials three types of algae were collected at the Jaunkemeri beach at the Gulf of Riga: brown, red and green algae. Brown algae *Fucusvesiculosus*, red algae and green algae *Furcellaria Cladophoraglomerata*, *C. rupestris*, *Enteromorpha* spp. are shown in Figures 1, 2 and 3.

Algae have been taken from the piles on the beach. Samples had very large admixture of sand, as well as strong odor. At first dry matter content was tested. For the red algae dry organic matter was extremely low, just 3.04 %. Therefore, additional pretreatment of algae was undertaken and sand with other inorganics was washed off and separated. However, still lot of sand was embedded in the algal pulp and was not fully separated.

For each sample delivered to the laboratory of the University of Life Sciences and Technologiethe chemical composition was determined according with ISO 6496: 1999. For each sample and inoculum dry matter, organic solids and ash content were determined. The analysis was made by standard methods [13; 14]. Similar methods are also used for energy crops by German researchers (Becker et. al. 2007) [12].



Fig. 1. Brown algae *Fucus vesiculosus*



Fig. 2. Red algae *Furcellaria*



Fig. 3. Green algae *Cladophora glomerata*, *C. rupestris*, *Enteromorpha* spp

Each group of raw materials was carefully weighed by weighing the sample and inoculum and afterwards thoroughly stirred. For all samples the same inoculum-digestate was used from the continuous operation bioreactor.

All algae prior to the analysis and filling in bioreactors were chopped, cut to 3 cm pieces. 0.75 liter bioreactors were filled with 20 g of raw material and 500 g inoculum (the weight recorded to 0.2 g accuracy). All data were recorded in the journal of experiments and computer. Bioreactors R2-R5 each was charged with a 20 g of brown seaweeds (BA), R6-R9 by 20g of red algae (RA), bioreactors R10-R13 with 20 g of green algae (GA) and bioreactors R14-R15 each containing 7 g BA, 7 g RA and 7g GA. All the bioreactors were connected to the gas storage bags with valves, placed in an oven and installed the operating temperature 38 ± 0.5 °C. Every day gas volume and composition were measured and recorded. Every day bioreactors were stirred in order to reduce the floating layer. Fermentation took place in a single filling mode and lasted until the biogas ceased to generate. Also digestate was weighed and dry matter, ash and organic dry matter content was determined.

Measurement accuracy was ± 0.02 pH, ± 0.025 L gas volume and ± 0.1 °C temperature. Periodically the composition of the produced biogas was determined, including CH₄, carbon dioxide CO₂, oxygen O₂ and hydrogen sulfide H₂S. Biogas production amount was investigated using laboratory equipment consisting of 16 0.75 liter bioreactors. Continuous operating temperature was provided by a SNOL type oven. Full dry matter was determined by dry weighing-machine Shimadzu at 105 °C temperature, composition of organic matter with the help of a drying oven Nabertherm and drying process was done with a special program at 550°C. The gas composition was measured with the gas analyzer GA 2000. The pressure was measured and calculated at the normal volume. The following table provides data calibrated to normal volume. Kern FKB 16KO2 scales was used for weight measurement, pH metering stationary device with accessories (PP-50).

Results and discussion

Extracted biogas and methane quantities were determined from all 16 digesters. Average values were calculated and presented in tables and visual graphs (Figures). Raw analytical results are shown in Table 1.

Table 1

Results of analyzes of raw material samples before anaerobic digestion

Bio-reactors	Raw material	pH	TS %	TS g	ASH, %	DOM, %	DOM, g	Weight, g
R1; R16	IN	7.57	4.2	21.0	19,48	80.52	16.909	500
R2-R5	BA	-	36.81	7.362	48.37	51.63	3.801	20
R2-R5	IN + BA	-	5.45	28.362	26.99	73.02	20.71	520
R 6 R 9	RA	-	29.54	5.908	40,34	59.66	3,525	20
R6-R9	IN + RA	-	5.17	26.909	24.06	75.94	20.434	520
R10-R13	GA	-	23.21	4.642	55.63	44.36	2,059	20
R10-R13	IN + GA	-	4.93	25.642	26.03	73.97	18.968	520
R14-R15	BA + RA + GA	-	29.85	6.27	47.59	52.41	3.286	21
R14-R15	IN + BA + RA + G A	-	5.23	27.27	35.94	74.06	20.195	521

Note: IN – inoculum; BA – brown algae; RA – red algae; GA – green algae; ASH – ashes; TS – total solids; DOM or dom – dry organic matter (on raw substrate Basis); R1-R16 – bioreactors.

As it is seen from the table, for all algae high dry matter and ash content were observed, but comparing to the traditional raw materials, very low dry organic matter content. This is explained by the large admixture of sand.

Biogas and methane amount extracted from each bioreactor is shown in Table 2. The average yield from starter yeast (inoculum) already deducted.

Table 2

Biogas and methane yields

Bioreactor/Raw material	Biogas, L	Biogas, L·g ⁻¹ dom	Methane, aver. %	Methane, L	Methane, L·g ⁻¹ dom
R1 500g IN	0.1	0.0059	3.00	0.003	0.00018
R16 500g IN	0.3	0.018	6.67	0.02	0.00118
Average R1, R16	0.2	0.012	5.78	0.012	0.0007
R2 500 g IN + 20g BA	0.9	0.237	14.56	0.131	0.034
R3 500g + 20g IN BA	1.1	0.289	18.27	0.201	0.053
R4 500 g IN + 20g BA	1.1	0.289	19.64	0.216	0.057
R5 500 g IN + 20g BA	1.1	0.289	12.91	0.142	0.037
Average R2- R5	1.05	0.276	16.48	0.173	0.046
± st.dev.	± 0.10	± 0.026	± 3.14	± 0.042	± 0.011
R6 500 g + 20 g IN RA	0.8	0.227	8.62	0.069	0.020
R7 500 g + 20 g IN RA	1.1	0.312	8.91	0.098	0.028
R8 500 g + 20 g IN RA	0.9	0.255	15.78	0.142	0.04
R9 500 g + 20 g IN RA	0.7	0.199	10.00	0.07	0.02
Average: R6-R9 IN + 20 g RA	0.875	0.248	10.86	0.095	0.027
± st.dev.	± 0.17	± 0.048	± 3.35	± 0.034	± 0.009
R10 500 g + 20 g IN GA	0.8	0.389	11.88	0.095	0.046
R11 500 g + 20 g IN GA	1.1	0.534	17.91	0.197	0.096
R12 500 g + 20 g IN GA	1.3	0.486	16.85	0.219	0.106
R13 500 g + 20 g IN GA	0.6	0.291	12.50	0.075	0.036
Average R10-R13 IN + 20 g GA	0.95 ± 0.31	0.425 ± 0.108	14.79 ± 3.04	0.360 ± 0.398	0.071 ± 0.035
± st.dev.					
R14 500g IN + 7g BA + 7gRA + 7gGA	0.9	0.274	16.44	0.148	0.045

Table 2 (continued)

Bioreactor/Raw material	Biogas, L	Biogas, L·g ⁻¹ dom	Methane, aver. %	Methane, L	Methane, L·g ⁻¹ dom
R15 500g IN + 7g BA + 7gRA + 7gGA	1.0	0.304	16.40	0.164	0.050
Average R14-R15 500g IN + 7g BA + 7gRA + 7gGA ± st.dev.	0.95 ± 0.07	0.289 ± 0.021	16.42 ± 0.03	0.156 ± 0.011	0.048 ± 0.004

Note: L·g⁻¹dom - liters per 1 g of dry organic Matter added (added fresh biomass into inoculums).

Average obtained biogas and methane quantities in comparison to other traditionally used feedstock for biogas production is very small. This is explained by the fact that algae are soaked with sea salts, which inhibit the anaerobic digestion process. This is shown by the very low methane content.

The results obtained in Ireland and Denmark [6-8] are more than three times better. This is explained by the fact that for the purpose of this study no special pretreatment of desalinization of the samples was undertaken to test biogas potential for mechanically collected and untreated feedstock with exemption of mechanical separation of inorganics so far as it is reasonable with simple washing.

The volume of biogas and methane from each bioreactor is shown in Figure 4.

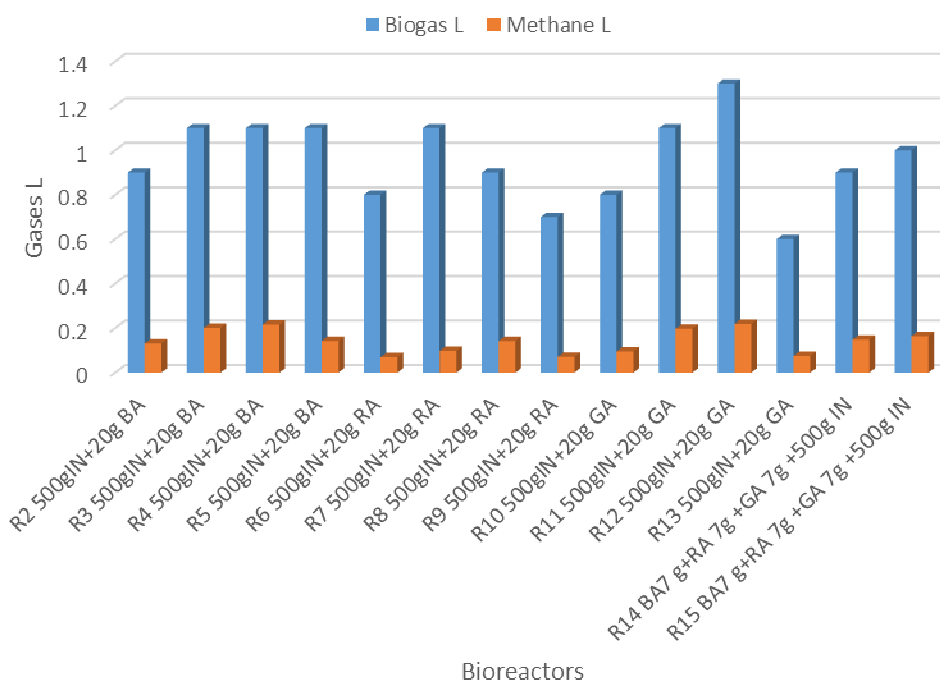


Fig. 4. Biogas and methane from each bioreactor

The biogas and methane specific outcome is shown in Figure 5. The results show that methane from all algae is representing a very small part of the biogas volume. The biogas composition shows larger amount of CO₂ present comparing to methane. This shows that the biomass hydrolysis took place, but methanogenesis was weak.

Thus, within raw materials methane forming bacteria was inhibited by presented of dissolved salts. The concentration of salt in particular samples was not part of this investigation taking into account the tested material close to the status of “as received” for the testing of algae biogas and methane potential with a view to apply anaerobic digestion for algae treatment and bio-energy generation.

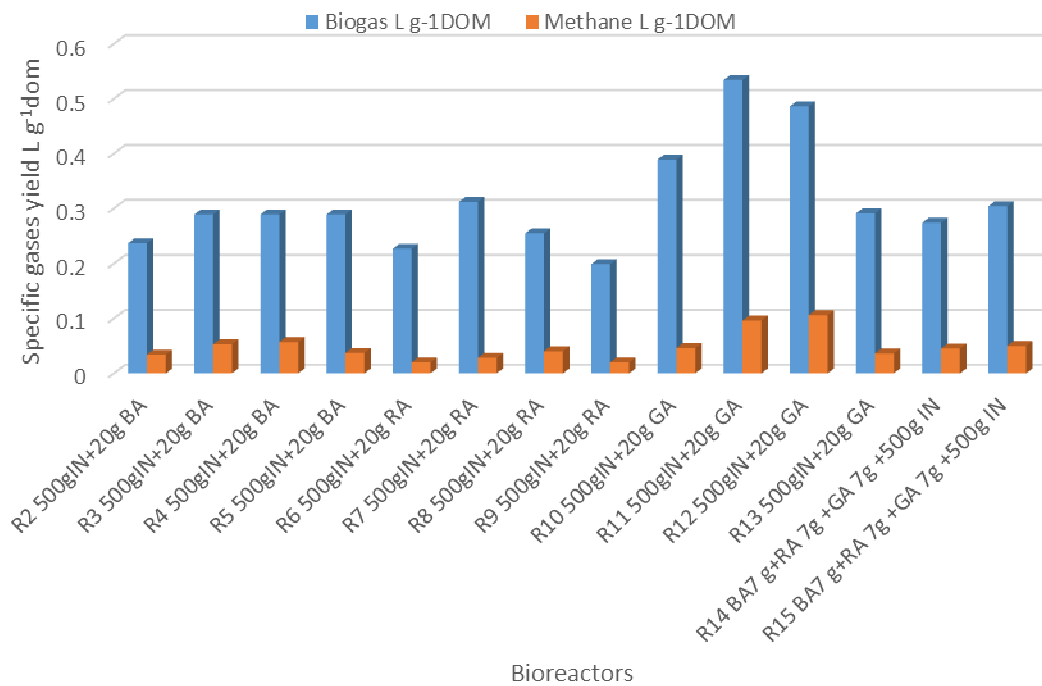


Fig. 5. Specific biogas and methane yields

Conclusions

1. Biogas (methane) retrieved from brown algae in average of 0.276 L·g⁻¹·dom (0.046 L·g⁻¹·dom) is a very low yield.
2. Biogas (methane) retrieved from red algae in an average of 0.248 L·g⁻¹·dom (0.027 L·g⁻¹·dom) is a very low yield.
3. Biogas (methane) retrieved from green algae in average of 0,425 L·g⁻¹·dom (0.071 L·g⁻¹·dom) is a slightly better yield, but still a poor result because of low methane content.
4. The overall results show that the algae washed out on the beach is a perspective bio-energy source, but with condition of additional pretreatment to lower the salt concentration. If the feedstock is used “as received” from beach clean-up, the methane yield will be too low to run economically feasible bio-energy generation.

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